

Genetically Modified Fuel-Ethanol Industrial Yeast Strains as Biocontrol Agents

Authors : Patrícia Branco, Catarina Prista, Helena Albergaria

Abstract : Industrial fuel-ethanol fermentations are carried out under non-sterile conditions, which favors the development of microbial contaminants, leading to huge economic losses. Wild yeasts such as *Brettanomyces bruxellensis* and lactic acid bacteria are the main contaminants of industrial bioethanol fermentation, affecting *Saccharomyces cerevisiae* performance and decreasing ethanol yields and productivity. In order to control microbial contaminations, the fuel-ethanol industry uses different treatments, including acid washing and antibiotics. However, these control measures carry environmental risks such as acid toxicity and the rise of antibiotic-resistant bacteria. Therefore, it is crucial to develop and apply less toxic and more environmentally friendly biocontrol methods. In the present study, an industrial fuel-ethanol starter, *S. cerevisiae* Ethanol-Red, was genetically modified to over-express AMPs with activity against fuel-ethanol microbial contaminants and evaluated regarding its biocontrol effect during mixed-culture alcoholic fermentations artificially contaminated with *B. bruxellensis*. To achieve this goal, *S. cerevisiae* Ethanol-Red strain was transformed with a plasmid containing the AMPs-codifying genes, i.e., partial sequences of TDH1 (925-963 bp) and TDH2/3 (925-963 bp) and a geneticin resistance marker. The biocontrol effect of those genetically modified strains was evaluated against *B. bruxellensis* and compared with the antagonistic effect exerted by the modified strain with an empty plasmid (without the AMPs-codifying genes) and the non-modified strain *S. cerevisiae* Ethanol-Red. For that purpose, mixed-culture alcoholic fermentations were performed in a synthetic must use the modified *S. cerevisiae* Ethanol-Red strains together with *B. bruxellensis*. Single-culture fermentations of *B. bruxellensis* strains were also performed as a negative control of the antagonistic effect exerted by *S. cerevisiae* strains. Results clearly showed an improved biocontrol effect of the genetically-modified strains against *B. bruxellensis* when compared with the modified Ethanol-Red strain with the empty plasmid (without the AMPs-codifying genes) and with the non-modified Ethanol-Red strain. In mixed-culture fermentation with the modified *S. cerevisiae* strain, *B. bruxellensis* culturability decreased from 5×10^4 CFU/mL on day-0 to less than 1 CFU/mL on day-10, while in single-culture *B. bruxellensis* increased its culturability from 6×10^4 to 1×10^6 CFU/mL in the first 6 days and kept this value until day-10. Besides, the modified Ethanol-Red strain exhibited an enhanced antagonistic effect against *B. bruxellensis* when compared with that induced by the non-modified Ethanol-Red strain. Indeed, culturability loss of *B. bruxellensis* after 10 days of fermentation with the modified Ethanol-Red strain was 98.7 and 100% higher than that occurred in fermentations performed with the non-modified Ethanol-Red and the empty-plasmid modified strain, respectively. Therefore, one can conclude that the *S. cerevisiae* genetically modified strain obtained in the present work may be a valuable solution for the mitigation of microbial contamination in fuel-ethanol fermentations, representing a much safer and environmentally friendly preservation strategy than the antimicrobial treatments (acid washing and antibiotics) currently applied in fuel-ethanol industry.

Keywords : antimicrobial peptides, fuel-ethanol microbial contaminations, fuel-ethanol fermentation, biocontrol agents, genetically-modified yeasts

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