

Association of G-174C Polymorphism of the Interleukin-6 Gene Promoter with Obesity in Iranian Population

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Abstract—Expression and secretion of inflammation markers are disturbed in obesity. Interleukin-6 reduces body fat mass. The common G-174C polymorphism in the promoter of IL-6 gene has been reported that effects on transcriptional regulation. The objective was to investigate association of the common polymorphism G-174C with obesity in Iranian population. The present study is cross sectional association study that included 242 individuals (110 men and 132 women). Serum IL-6 levels, C-reactive protein, fasting blood glucose and blood lipids profile were measured. BMI and WHR were calculated. Genotyping is carried out by PCR and RFLP. The frequencies of G and C allele were 64.5% and 35.5%, respectively. The G-174C polymorphism was not associated with BMI and WHR. However in obese individual, fasting blood glucose was significantly higher in carrier of C allele compared with the non-carrier. The IL-6 G-174C polymorphism is not a risk factor for obesity in Iranian population.

Keywords—Interleukin 6, Polymorphism genetic, Obesity.

I. INTRODUCTION

THE prevalence of obesity has been increasing rapidly in most of the world [1]. Numerous studies indicate that excess body fat induce a multitude of co-morbid conditions such as diabetes, cardiovascular disease metabolic syndrome and cancer [2]. The importance of genetics factors to control of human body weight and composition, in concert with environmental effects has been well established. It has been reported that chronic low-grade activation of the immune system plays a central role in the etiology of obesity [2] [3]. Obesity can be expressed as an inflammation condition [1]. In the obesity state the expression and secretion of inflammation markers are disturbed. So, one of the best ways for identification of body fat regulative mechanisms has been exploration of the genetic relationship between pro-inflammation cytokines and adiposity [2]. IL-6, a major pro-inflammation cytokine expressed in several tissues such as, adipose tissue, muscles, immune cells and hypothalamus are associated with regulatory of energy balance in human [3]. High circulation and adipose tissue levels of IL-6 have been correlated with obesity and visceral fat [4]. Several SNPs

(Single nucleotide polymorphism) have been identified in the IL-6 gene. The common G-174C polymorphism in the promoter of IL-6 gene has been reported that affects on transcriptional regulation [2], [3]. Several studies suggest that -174 G-containing haplotypes are stronger enhancer of IL-6 transcription than those containing C allele [5]. Whereas, other studies observed that polymorphism G-174C was not associated with obesity and IL-6 levels. However, studies on the effects this polymorphism on circulating IL-6 and gain weight generated conflicting result.

Based on these finding, we examined the association of the common polymorphism G-174C (rs1800795) with obesity in obese and non-obese from prospective cohort.

II. METHODS

A. Study subject

The sample for the present study included 242 individuals (110 men and 132 women) overall, who were selected for the study from the Tehran Lipid and Glucose Study (TLGS) cohort. Peripheral blood, blood clot and blood with anti-coagulation EDTA were obtained. Hip and waist circumference and hypertension were measured. Body Mass Index (BMI) was calculated with the formula: weight (kg)/height (m²). Waist to hip ratio (WHR) was calculated as waist circumference divided by hip circumference. BMI ≥ 30 and WHR (> 0.85 for women; > 1.0 for men) were used as indices of obesity.

B. Biochemical analyses

The serum concentration of triglyceride (TG), total cholesterol and glucose were measured by commercial kits (Pars Azmoun, Tehran, Iran). The HDL-C of serum was determined by apolipoprotein sediment with phosphotangenic acid. LDL-C in subjects with TG < 400 mg/dl was assayed used of the Friedewald formula. The serum IL-6 and high sensitive CRP (hsCRP) were measured by using

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an ELISA kits (Diacclone, Besancon, France and dbc, Ontario, Canada) respectively.

C. DNA isolation and genotyping:

Genomic DNAs were obtained from peripheral blood leukocytes by salting out method. The polymorphism of the IL-6 promoter region at -174 was studied by PCR-RFLP. The region of interest was amplified by PCR using primers 5-TGACTTCAGCTTTACTTTGT-3 and 5-AATCTTAATAAGGTTTCCA-3. The reaction was carried out in a final volume of 25 μ L containing 1.5 mmol/L of MgCl₂, 0.2 mmol/L of each dNTP, 0.2 mmol/L of each primer and 1 unit of Taq polymerase (Cinagene Co, Tehran, Iran). DNA was amplified during 30 cycles with an initial denaturation of 10 minutes at 94°C and a final extension of 10 m at 72°C. The cycle program consisted in 1 minute denaturation at 94°C, 1 minute and 35s annealing at 62°C and 1 m extension at 72°C. The PCR product was digested by adding 10U of Nla III restriction enzyme at 37°C overnight, separated the digested fragments by Agarose Gel Electrophoresis and detected by ethidium bromide staining. The polymorphism is due to a replacement of G by C at position -174. The identified genotypes were named according to the presence or absence of the enzyme restriction sites, so Nla III (GG), (GC) and (CC) are homozygote for the presence of the site (153/40 bp), and heterozygote for the presence/absence of the site (193/153/40) and homozygote for the absence of the site (198 bp), respectively.

D. Statistical analysis:

To analysis data using the SPSS program (version 16.0). To compare of variables between the three different genotypes and carriers of G-174C polymorphism respectively, we used the Kruskal-Wallis (K-W one-way) and Mann-Whitney due to non-normally distributed data. X² analysis was used for the investigation of the presence of Hardy-Weinberg equilibrium. Results were expressed as means \pm SE.

III. RESULT

In this study, the genotypes were in Hardy-Weinberg equilibrium. The frequencies of G and C allele in the G-174C IL-6 polymorphism were 64.5% and 35.5%, respectively. In all, 156 subjects(64.5%) had a GG genotype, 71subjects (30.3%) a GC genotype and 15 subjects(6.2%) had a CC genotype. all of the groups(men , women and obese, non-obese) frequency of carrying G allele were more than carrying C allele exception in obese men that frequency of C allele was more than G allele (Table I). The relation between G-174C genotype and various vital, blood chemistry parameters and geometric of adiposity measures (MBI, Waist circumference and Waist to hip ratio) is shown in table II. The G-174C polymorphism was not associated with difference in age, blood pressure, fasting blood glucose, total cholesterol, LDL, HDL, triglycerides, serum IL-6, hsCRP. Moreover, the G-174C polymorphism was not associated with BMI and WHR. However in obese individual, fasting glucose blood

was significantly higher in carrier of allele C compared with the non-carrier ($P=0.04$) (Table III). There was a tendency in obese individuals that carrying C allele has higher BMI, WHR and weight than carrying G allele and has lower IL-6 concentration (Table 2).

IV. DISCUSSION

The present study shows that the IL-6 G-174C polymorphism is not significantly associated with increased BMI in Iranian population. Several studies examined the associations between the common polymorphism in the IL-6 gene promoter in human. Some of these studies expressed that, the IL-6 polymorphism G-174C have been related with obesity especially central adiposity [5], [6]. Some surveys have reported that meaningful relation between IL-6 G-174C gene polymorphism and adiposity were not observed [7], [8]. Some other studies have reported that IL-6 increase energy expenditure in rodents and human, suppresses body fat and limits late-onset obesity [9]. The C allele appears to be related with low concentration of IL-6 in variant cell systems and decrease body temperature and energy expenditure [10], [11], [12]. Decrease of IL-6 concentration and its association with the G-17C allele has been reported in some studies [11], [13], not all of them [14], [15]. In this study, we observed that obese individual carrying the G allele had higher IL-6 concentration than the C allele carrier obese individuals but this difference was not significant. Circulating IL-6 is derived from different cell types such as adipose tissue, muscle cells and hypothalamus, that all of them contribute to the control of energy balance [4]. In addition, there are considerable irregular variations of circulating IL-6 levels in during the day [16]. In other hand, genetically determined decrease in IL-6 production per weight adipose tissue probably in theory cause increased adiposity [17]. The high concentration of IL-6 in obese individuals could involve resistant of IL-6 in a like path as they are patterned to be Leptin resistant [18].

Nevertheless, evidence have been shown that the effects of IL-6 on adipose tissue and energy expenditure are exerted at the level the central nervous system (CNS) instead of the periphery [9], [19], [20]. As well as, the IL-6 level in the CNS seem to be controlled in a various way than circulating levels of IL-6 [21], raising the possibility level of circulating IL-6 do not return the anti-obesity potential of IL-6 [3]. Obesity, insulin resistant and metabolic syndrome seems to be related with serum IL-6 levels [22]. In 1990, it was reported that treatment with high amount of IL-6 increase insulin resistant [23], [24]. However, some more studies have not shown that after IL-6 treatment any increase in blood glucose [25] [26]. Interestingly, anti-IL-6 receptor antibody treatment has been shown to cause increased level of blood glucose in some patient with rheumatoid arthritis [27]. In the present study, we observed that fasting blood glucose in obese individuals carrying C allele was higher than obese individuals carrying G allele. So, the effect of IL-6 appears complex and both increased and decreased activity of IL-6 cause enhanced insulin resistant. In conclusion, as indicated in

the study, the C allele is probably not affecting gain weight in Iranian population, maybe due to interaction with unknown environmental and genetic factors.

REFERENCES

[1] Hans Hauner, Symposium on 'Biology of obesity' Secretory factors from human adipose tissue and their functional role, Proceedings of the Nutrition Society , pp163–169, 2005.

[2] Lu Qi, Cuilin Zhang, Rob M. van Dam and Frank B. Hu, Interleukin-6 Genetic Variability and Adiposity: Associations in Two Prospective Cohorts and Systematic Review in 26,944 Individuals, Journal of Clinical Endocrinology & Metabolism, 92: 93618-3625, 2007.

[3] I Wernstedt, A-L Eriksson, A Berndtsson, J Hoffstedt, S Skrtic, T Hedner, LM Hulthe, O Wiklund, C Ohlsson and J-O Jansson , A common polymorphism in the interleukin-6 gene promoter is associated with overweight, International Journal of Obesity 28, pp 1272–1279, 2004.

[4] Victoria Rotter Sopasakis, Madeleine Sandqvist, Birgit Gustafson, Ann Hammarstedt, Martin Schmelz, Xiaolin Yang, Per-Anders Jansson, and Ulf Smith , High Local Concentrations and Effect on Differentiation Implicate Interleukin-6 as a Paracrine Regulator, Obesity res. 12 :3(454-461), 2004.

[5] Berthier MT, Paradis AM, Tcherno A, Bergeron J, Prud'homme D, Despres JP, Voh I, MC The interleukin 6–174G/C polymorphism is associated with indices of obesity in men. J Hum Genet 48:14–19, 2003.

[6] Klipstein-Grobusch K, Mohlig M, Spranger J, Hoffmann K, Rodrigues FU, Sharma AM, Klaus S, Pfeiffer AF, Boeing H, Interleukin-6 g-174G>C promoter polymorphism is associated with obesity in the EPIC-Potsdam Study. Obesity (Silver Spring) 14:14–18, 2006.

[7] Lieb W, Pavlik R, Erdmann J, Mayer B, Holmer SR, Fischer M, Baessler A, Hengstenberg C, Loewel H, Doering A, Riegger GA, Schunkert H, No association of interleukin-6 gene polymorphism (-174 G/C) with myocardial infarction or traditional cardiovascular risk factors. Int J Cardiol 97:205–212, 2004.

[8] Moffett SP, Zmuda JM, Cauley JA, Stone KL, Nevitt MC, Ensrud KE, Hillier TA, Hochberg MC, Joslyn G, Morin P, Cummings, Association of the G-174C variant in the interleukin-6 promoter region with bone loss and fracture risk in older women. J Bone Miner Res 19:1612–1618, 2004.

[9] Wallenius V, Wallenius K, Ahren B, Rudling M, Carlsten H, Dickson SL, Ohlsson C, Jansson J-O, Interleukin-6-deficient mice develop mature-onset obesity. Nat Med 8: 75–79, 2002.

[10] Fishman D, Faulds G, Jeffery R, Mohamed-Ali V, Yudkin JS, Humphries S, Woo P. The effect of novel polymorphisms in the interleukin-6 (IL-6) gene on IL-6 transcription and plasma IL-6 levels, and an association with systemic-onset juvenile chronic arthritis. J Clin Invest; 102: 1369–1376, 1998.

[11] Terry CF, Loukaci V, Green FR. Cooperative influence of genetic polymorphisms on interleukin 6 transcriptional regulation. J Biol Chem; 275:18138–18144, 2000.

[12] Acalovschi D, Wiest T, Hartmann M, Farahmi M, Mansmann U, Auffarth GU, Grau AJ, Green FR, Grond-Ginsbach C, Schwanninger M. Multiple levels of regulation of the interleukin-6 system in stroke. Stroke; 34: 1864–1869, 2003.

[13] Kazumi T, Kawaguchi A, Hirano T, Yoshino G. C-reactive protein in young, apparently healthy men: associations with serum leptin, QTC interval, and high-density lipoprotein-cholesterol. Metabolism, 52: 1113–1116, 2003.

[14] Kubaszek A, Pihlajamaki J, Punnonen K, Karhapaa P, Vauhkonen I, Laakso M. The C-174G promoter polymorphism of the IL-6 gene affects energy expenditure and insulin sensitivity. Diabetes, 52: 558–561, 2003.

[15] Brull DJ, Montgomery HE, Sanders J, Dhamrait S, Luong L, Rumley A, Lowe GD, Humphries SE. Interleukin-6 gene -174g4c and -572g4c promoter polymorphisms are strong predictors of plasma interleukin-6 levels after coronary artery bypass surgery. Arterioscler Thromb Vasc Biol; 21: 1458–1463, 2001.

[16] Sothorn RB, Roitman-Johnson B, Kanabrocki EL, Yager JG, Roodell MM, Weatherbee JA, Young MR, Nenchausky BM, Scheving LE. Circadian characteristics of circulating interleukin-6 in men. J Allergy Clin Immunol; 95: 1029–1035, 1995.

[17] Farooqi IS, Keogh JM, Kamath S, Jones S, Gibson WT, Trussell R, Jebb SA, Lip GY, O'Rahilly S. Partial leptin deficiency and human adiposity. Nature; 414: 34–35, 2001.

[18] Flier JS. Obesity wars: molecular progress confronts an expanding epidemic. Cell 2004; 116: 337–350.

[19] Wallenius K, Wallenius VW, Sunter D, Dickson SL, Jansson J-O. Intracerebroventricular interleukin-6 treatment decreases bodyfat in rats. Biochem Biophys Res Comm; 293: 560–565, 2002.

[20] Rothwell NJ, Busbridge NJ, Lefevre RA, Hardwick AJ, Gaudie J, Hopkins SJ. Interleukin-6 is a centrally acting endogenous pyrogen in the rat. Can J Physiol Pharmacol; 69: 1465–1469, 1991.

[21] Stenlof K, Wernstedt I, Fjallman T, Wallenius V, Wallenius K, Jansson JO. Interleukin-6 levels in the central nervous system are negatively correlated with fat mass in overweight/obese subjects. J Clin Endocrinol Metab; 88: 4379–4383, 2003.

[22] Fernandez-Real JM, Ricart W. Insulin resistance and chronic cardiovascular inflammatory syndrome. Endocr Rev; 24: 278–301, 2003.

[23] Tsigos C, Papanicolaou DA, Kyrou I, Defensor R, Mitsiadis CS, Chrousos GP. Dose-dependent effects of recombinant human interleukin-6 on glucose regulation. J Clin Endocrinol Metab; 82: 4167–4170, 1997.

[24] Stouthard JM, Romijn JA, Van der Poll T, Endert E, Klein S, Bakker PJ, Veenhof CH, Sauerwein HP. Endocrinologic and metabolic effects of interleukin-6 in humans. Am J Physiol; 268: E813–E819, 1995.

[25] Lyngso D, Simonsen L, Bulow J. Metabolic effects of interleukin-6 in human splanchnic and adipose tissue. J Physiol; 543: 379–386, 2002.

[26] Steensberg A, Fischer CP, Sacchetti M, Keller C, Osada T, Schjerling P, van Hall G, Febbraio MA, Pedersen BK. Acute interleukin-6 administration does not impair muscle glucose uptake or whole-body glucose disposal in healthy humans. J Physiol; 548: 631–638, 2003.

[27] Nishimoto N, Yoshizaki K, Maeda K, Kuritani T, Deguchi H, Sato B, Imai N, Suemura M, Kakehi T, Takagi N, Kishimoto T. Toxicity, pharmacokinetics, and dose-finding study of repetitive treatment with the humanized anti-interleukin 6 receptor antibody MRA in rheumatoid arthritis. Phase I/II clinical study. J Rheumatol; 30: 1426–1435, 2003.

TABLE 1
 FREQUENCY OF G AND C ALLELE

	Female		Male		Total
	obese	Non-obese	obese	Non-obese	
Allele G	15(38/4)	40(72/2)	25(64.1)	76(69.7)	156(64.5)
Allele C	24(61.6)	15(27/7)	14(35.9)	33(30.3)	86(35.5)

Data are shown as no (%).

TABLE II
 CLINICAL AND BIOCHEMICAL CHARACTERISTICS ACCORDING TO THE G-174C GENOTYPE

	-174 IL-6 genotype			Pvalue (K-W)
	GG	GC	CC	
Subject(No. /%)	156/64	71/29	15/6	
Age	38.43±19.06	34.58±16.99	42.20±14.07	
BMI	25.28±15.24	25.39±4.91	26.57±6.60	0.17
WHR	0.86±0.08	0.86±0.09	0.85±0.08	0.95
Weight	64.66±15.24	65.94±13.98	69.70±13.57	0.50
Ln SBP(mmHg)	4.73±0.13	4.70±0.13	4.70±0.12	0.22
DBP(mmHg)	73.21±8.13	72.49±8.71	73.73±8.74	0.75
Fb-glucose	88.5±28.75	87.50±55.86	91.00±20.76	0.11
Total cholesterol(mM)	191.58±41.47	186.90±42.97	193.33±43.99	0.62
LDL(mM)	120.79±34.95	116.34±34.08	120.28±41.14	0.65
HDL(mM)	43.12±9.94	42.14±9.54	45.86±11.31	0.54
LnTriglycerides (mM)	4.82±0.44	4.79±0.58	4.78±0.52	0.65
IL-6(pg/ml)	1.49±0.97	1.48±0.99	1.71±0.68	0.55
hsCRP(ng/ml)	6.73±1.48	6.50±1.56	6.32±1.91	0.53

SBP=systolic blood pressure, DBP=diastolic blood pressure, Fb-glucose=fasting blood glucose, IL-6=interleukin 6, hsCRP=high sensitive C-reactive protein. The *p*-value is for comparison of the three genotypes (Kruskal -Wallis). Data are shown as mean ± SEM

TABLE III
 CLINICAL AND BIOCHEMICAL CHARACTERISTICS ACCORDING TO ALLELE CARRIERS IN G-174C SNP

Carrier	obese		p-value	Non-obese		p-value
	G	C		G	C	
BMI	31.53±2.78	32.49±4.09	0.63	23.97±2.11	23.90±2.91	0.84
Weight	75.57±8.16	80.74±11.49	0.15	62.37±15.41	63.18±12.29	0.65
WHR	0.89±0.05	0.92±0.07	0.20	0.85±0.09	0.84±0.08	0.88
Fb-glucose (mg/dl)	89.5±14.9	95.00±33.4	0.04	88.5±30.8	88.5±30.8	0.05

The *p* -value are for comparison of the carriers (Mann-Whitney). Data are shown as mean ± SEM