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Study on the effect of Sulphur, Glucose, Nitrogen and plant residues on the Immobilization of Sulphate-S in Soil

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***Abstract*—**In order to evaluate the relationship between the sulphur (S), glucose (G), nitrogen (N) and plant residues (st), sulphur immobilization and microbial transformation were monitored in five soil samples from 0-30 cm of Bastam farmers fields of Shahrood area following 11 treatments with different levels of Sulphur (S), glucose (G), N and plant residues (wheat straw) in a randomized block design with three replications and incubated over 20, 45 and 60 days, the immobilization of SO_4^{-2}-S presented as a percentage of that added, was inversely related to its addition rate. Additions of glucose and plant residues increased with the C-to-S ratio of the added amendments, irrespective of their origins (glucose and plant residues). In the presence of C sources (glucose or plant residues). N significantly increased the immobilization of SO_4^{-2}-S, whilst the effect of N was insignificant in the absence of a C amendment. In first few days the amounts of added SO_4^{-2}-S immobilized were linearly correlated with the amounts of added S recovered in the soil microbial biomass. With further incubation the proportions of immobilized SO_4^{-2}-S remaining as biomass-S decreased. Decrease in biomass-S was thought to be due to the conversion of biomass-S into soil organic-S. Glucose addition increased the immobilization (microbial utilization and incorporation into the soil organic matter) of native soil SO_4^{-2}-S. However, N addition enhance the mineralization of soil organic-S, increasing the concentration of SO_4^{-2}-S in soil

***Keywords*—**Immobilization, microbial biomass, sulphur, nitrogen, glucose.

I. Introduction

SULPHUR (S) immobilization in soil is the process through which mineral S is incorporated into soil organic compounds [10]. This process together with the mineralization of organic-S, regulates the accumulation and cycling of S in the soil and affects S availability to plants [6]. This immobilization is believed to be microbially mediated which includes the conversion of S into the microbial biomass (microbial utilization). However, the dynamics of the immobilization process and the mechanisms through which immobilization is associated with microbial transformations [2], (the utilization and turnover of S) remain poorly understood. Recent developments in methodologies for measuring soil microbial biomass-S [10] have facilitated progress towards the quantification of S transfer (rate and magnitude) between SO_4^{-2}-S microbial biomass-S and organic-S pools in soil. [9] have shown that S immobilization in soils amended with plant residues (barley straw or rape leaves) is highly correlated with increases in microbial biomass-C. In a separate study using six soils with different properties, it was shown that the amounts of added SO_4^{-2}-S immobilized were comparable with those converted into the microbial biomass over the first few days [6], [7]. The subsequent incorporation of the immobilized SO_4^{-2}-S into the soil organic matter was through to depend on the turnover of the soil microbial biomass. Furthermore, this study demonstrated that the immobilization rates of SO_4^{-2}-S were influenced by soil properties such as clay and organic matter contents, the size of the microbial biomass and the available S. However, the extent to which these factors interact and affect the immobilization rates of S remained unknown.

This study showed the dynamics of the immobilization of SO_4^{-2}-S using amendments containing different rates of SO_4^{-2}-S, different form of carbon (glucose, straw) and N nutrient. The initial Immobilization rates of added SO_4^{-2}-S were correlated with the amounts converted to soil microbial biomass-S over first few days. Properties of decomposed biomass-S converted into the soil organic-S (incorporated into the soil organic matter) were determined on further incubation. Our objective was to establish the quantitative relationship between the immobilization and microbial transformation of S and to estimate the effects of factors such as the amounts of SO_4^{-2}-S, the supply of C and N on the immobilization and availability of SO_4^{-2}-S in soil. These data are essential in proving the hypothesis that the incorporation of SO_4^{-2}-S into the soil organic matter via the microbial biomass is the primary mechanism for the immobilization of inorganic-S in soils [6], [1].

II. Materials And Methods

Composite samples from 0-30 cm depth of the five farmer's field were collected, air dried, and passed through 2 mm sieve and kept for 10 days in 25°C and 100% humidity. Soil physical and chemical properties were done by standard methods and are presented in Table I.

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TABLE I
PHYSICAL AND CHEMICAL PROPERTIES OF THE SOILS

Site	Clay (%)	O.C (%)	pH	Ec (mS cm^{-1})	Total S mg kg^{-1}	Available. S mg kg^{-1}
1	33	0/88	7.65	0.74	278	1.4
2	42	1.05	7.44	0.59	311	3.4
3	31	0.79	7.26	0.61	252	3.2
4	37	0.74	7.64	0.83	195	2.6
5	44	0.88	7.48	0.79	205	3.6

Sixty gram of dried soil were weighed into 125 ml jars and amended with S, N, glucose and plant residues as detailed in table 2. Controls were left unamended. Sulphur as K_2SO_4 (S_{10} and S_{25} µ g g^{-1}), N as KNO_3 and appropriate amounts of glucose and wheat straw were added to soil. For each treatment, a solution was prepared by dissolving the appropriate amounts of glucose, KNO_3 and K_2SO_4 in 100 ml distilled water. An aliquot of the solution (4 ml) was mixed with each soil portion.

Before the addition of the solutions, plant residues (0.3 g) which had been dried (35° C) and ground were mixed with the soil portions as required. The control soil was treated with 4 ml distilled water to maintain equivalent moisture content to that of the amended soils. Following amendment, the soils were placed in 2.5 glass bottles, sealed and kept at 25° C and 100% humidity.

TABLE II
DESCRIPTION OF THE TREATMENTS

Treatments	SO_4^{-2}-S (µg g^{-1} soil)	NO_3^-N (µg g^{-1} soil)	Glucose (µg g^{-1} soil)	Straw-C (µg g^{-1} soil)
Control	---	---	---	---
S_{10}	10	---	---	---
S_{25}	25	---	---	---
S_{10}+N	10	100	---	---
S_{25}+N	25	100	---	---
S_{10}+G	10	---	2500	---
S_{25}+G	25	---	2500	---
S_{10}+G+N	10	100	2500	---
S_{25}+G+N	25	100	2500	---
S_{10}+St+N	10	100	---	2500
S_{25}+St+N	25	100	---	2500

The contents of SO_4^{-2}-S and the microbial biomass-S were determined after 20, 45 and 60 days of incubation. At each sampling, one portion of soil from each treatment was removed and subdivided by weighing (6×10g) into centrifuge tubes (45 ml). Three of the sub-samples were fumigated for 24 h in $CHCl_3$ vapor [10]. The remaining three sub-samples were used as the controls and left unfumigated. All of the sub-samples were extracted in 10 mM $CaCl_2$ (20 ml) by shaking for 60 min at 400 rev min^{-1} on an end-over-end shaker. Extracts were filtered through Whatman No. 42 filter paper and stored at –18°C prior to analysis.

Soil microbial biomass-S was measured by the procedures described by [10]. Briefly, total extractable-S was determined using [5] following conversion of organic-S in the extract into SO_4^{-2}-S. This was done by digesting an aliquot of the extracts (5 ml) in a 10 ml graduated glass tube for 24 h in a sand bath (160°C), using H_2O_2 (AR grade, 30% v/v, 1.5 ml). Total biomass-S was calculated from the relationship $B_s = F_s/K_s$, where Fs is the difference between total extractable-S in the fumigated soil and that in the control soil, K_s with the conversion factor (0.31), determined by [10]. Analysis of variance (ANOVA) was performed on all data sets. Data from all treatment were combined in correlation and regtession analysis. The statistical package Minitab12 and excel were used.

III. RESULTS AND DISCUSSION

Immobilization of added sulphate-S

The amounts of added SO_4^{-2}-S recovered from all of the treatments using 10 mM CaCl2 decreased over 60 days incubation (Fig. 1). These decreased were similar in that they were initially rapid but became slow with extended incubation. This agrees with the findings of [6] who measured the immobilization of added SO_4^{-2}-S in a number of soils with different properties. In this study results showed that decreases in the recovery of SO_4^{-2}-S were greater in those soils receiving larger additions of amendments (S_{25}, S_{25} + G and S_{25}+ G + N treatments, compared with S_{10}, S_{10}+G and S_{10}+G +N treatments; Table II). This suggests that the amount of SO_4^{-2}-S immobilized (converted into soil microbial biomass-S or incorporated into soil organic matter) in soil was, as expected, positively correlated with the addition rate. However, the extent to which SO_4^{-2}-S was immobilized, as a percentage of addition, was inversely correlated to the addition rate. This is indicated clearly by the fact that the percentage of SO_4^{-2}-S immobilized was smaller in those soils receiving the larger additions of SO_4^{-2}-S.

In this experiment treatments with the addition of glucose markedly increased the immobilization of SO_4^{-2}-S, particularly over the first 10 days of incubation. For example, in treatment S_{10} + G + N the recoveries of SO_4^{-2}-S in first 10 days was 50%, and by the end of incubation immobilization had decreased to over 35%.

In contrast, in those treatments receiving no additional C source (S_{10}+S_{10}+N), the immobilization of SO_4^{-2}-S was less that 15% throughout the incubation. These results were expected, as it has been shown that the addition of labial substrate such as glucose can results in a rapid increase in microbial biomass which requires mores nutrient from the soil [4], [9], [6].

Based on the comparison of treatments S_{10} and S_{10} + N, and S_{25} and S_{25} + N, the addition of N without added C was unlikely to change significantly the immobilization of SO_4^{-2}-S (Fig.2). However, the combination of N with glucose was shown to have a positive effect on S immobilization in soil. As shown in Fig. 2, in the treatments of S_{10} + G + N and S_{25} + G + N, the immobilization of SO_4^{-2}-S increased by 12- 15% (as a percentage of the addition rates) by 20 days of incubation, when compared with treatments S_{10} + G and S_{25} + G. Thus, the supply of N can limit S immobilization in soil, particularly during rapid growth of the microbial biomass, as found following glucose addition [8].

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Additions of straw residues (S_{25} + St + N) increased the immobilization of SO_4^{-2}-S, compared with the treatment S_{25} + N which provide an equivalent amount of SO_4^{-2}-S and N but contained no residue amendment (Fig. 2). This was presumably a result of the rapid growth of the soil microbial biomass following the addition of plant residues [7], [1], [9]. However, the effect of plant residues on the immobilization of SO_4^{-2}-S was much smaller than that of glucose, since the amounts of SO_4^{-2}-S immobilized in treatments S_{25} + St +N were apparently less than those found in treatment S_{25} + G + N. Straw is considerably less labile and, at the same addition rate, might be expected to produce smaller increase in soil microbial biomass and S immobilization than either glucose or the straw residues [7], [9]. Previous studies by researchers have shown that the incorporation of plant residues with narrow C-to-s ratio (<200:1) can increase the contents of SO_4^{-2}-S in the soil, whereas the incorporation of those plant residues which have wide C-to-S ratios (<400:1) may result in a net immobilization of soil inorganic-S [7], [9]. It can be conclude that there are close relationships between the immobilization of SO_4^{-2}-S and the microbial utilization and turnover of S. The data support the hypothesis that primary mechanism by which SO_4^{-2}-S is incorporated into the soil organic matter is via the microbial biomass [6]. Since immobilization is dependent upon microbial utilization and turnover, the extent to which available-S (SO_4^{-2}-S) can be immobilized is determined by both the amount of available-S and the availability of an utilizable C source.

REFERENCES

[1] S.Bhupinderpal., Z. Rengel and J.W. Bowden, Carbon, nitrogen and sulphur cycling following incorporation of canola residue of different sizes into a nutrient-poor sandy soil. Biology and Biochemistry., Vol. 38, pp.32-42. 2006.

[2] H.Chowdhury., K.M. Kouno, T. AndoCritical sulphur concentration and sulphur requirement of microbial biomass in a glucose and cellulose-amemded regosol. Biol. Fertil. Soils ., Vol. 32, pp. 310-317. 2000.

[3] E.Eriksen, Gross sulphur mineralization–immobilization turnover in soil amended with plant residues. Soil biology and Biochemistry. Vol. 37, pp. 2216-2224. 2005.

[4] A.Ghani, R.G. McLarn, and R.S. Swift, The incorporation and transformations of 35S in soil: Effects of soil conditioning and glucose or sulphate additions. Soil Biol. Biochem. Vol. 2, pp. 327-335 .1993.

[5] Massoumi, A. and A. H. Cornfield, A rapid method for determinating sulphate in water extracts of soils. Analyst, London. Vol. 88, pp. 321-322. 1963.

[6] A.G.O'Donnell., J. Wu, and J.K. Syers, Sulphate-S amendments in soil and their effects on the transformation of soil sulphur.Soil Biology and Biochemistry. Vol 26, pp. 1507-1514. 1994.

[7] T.Pennanen, Caul, S., Daniell, J.J., Griffiths, B.S., Ritz, K, and R.E., Wheatley, Community-level responses of metabolically active soil micro-organism to the quantity and quality of substrate inputs. Soil Biol. Biochem. Vol. 36, pp. 841-848 .2004.

[8] Vong, P.C. S., Piutti, S.S Deschaumes, E, Benizri and A. Guckert, Sulphur immobilization and arylsulphatase activity in two calcareous arable and fallow soils as affected by glucose additions. Geoderma.Vol. 148, pp. 79-84 .2008.

[9] J.Wu, AG O'Donnell, JK SyersMicrobial growth and sulphur immobilization following the incorporation of plant residues into soil. Soil Biol. Bioch ., Vol. 25, pp.1567-1573. 1993.

[10] J.Wu, D. O' Donnell, A.G, Z. L. He, and J. K. Syers, Fumigation-extraction method for the measurement of soil microbial biomass-S. Soil Biol. Biochem.,Vol. 26: pp 117-125. 1994.

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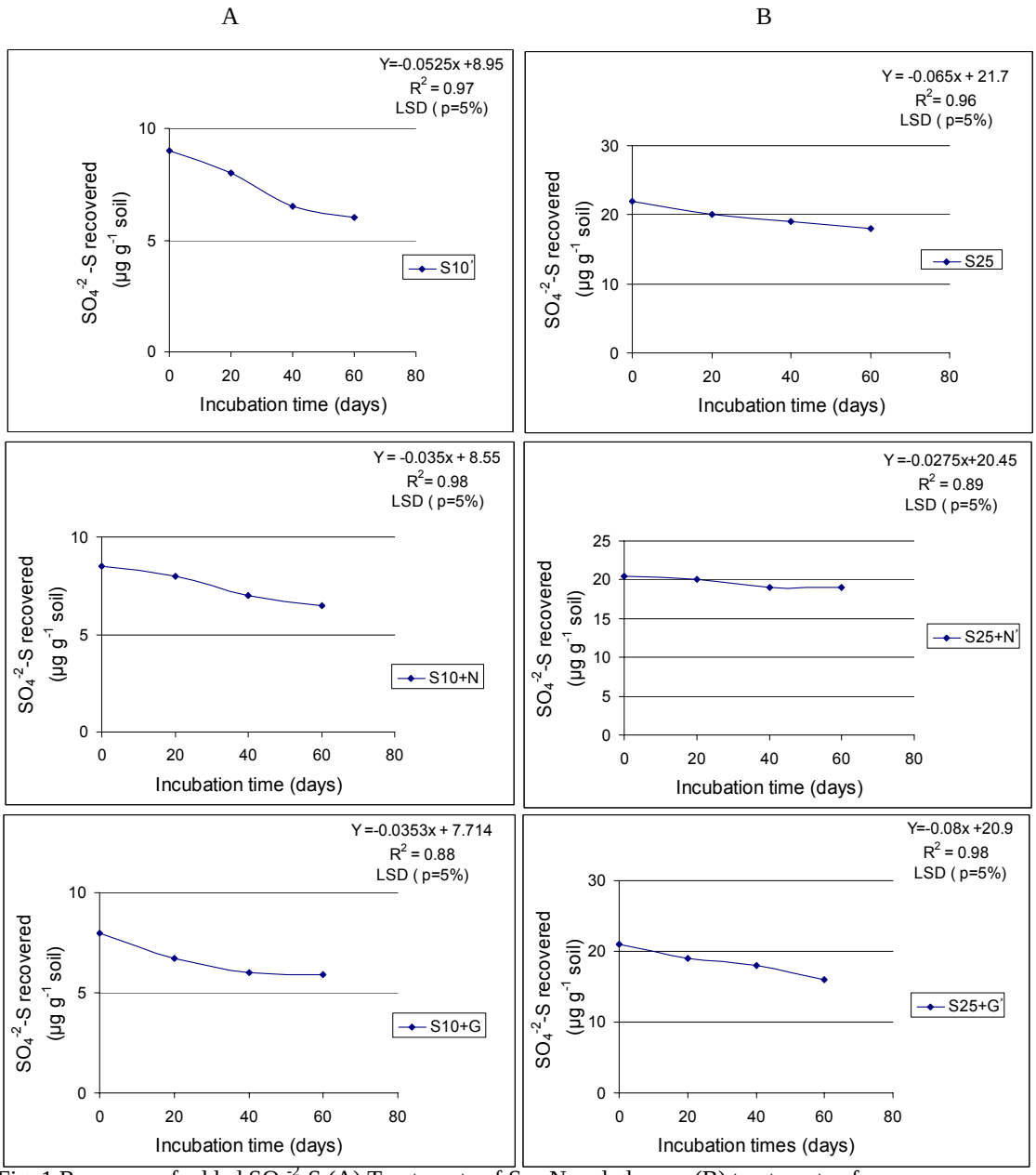


Fig. 1 Recovery of added SO_4^{-2}-S (A) Treatments of S_{10}, N and glucose (B) treatments of S_{25}, N, and glucose. Details of the treatments are described in Table II

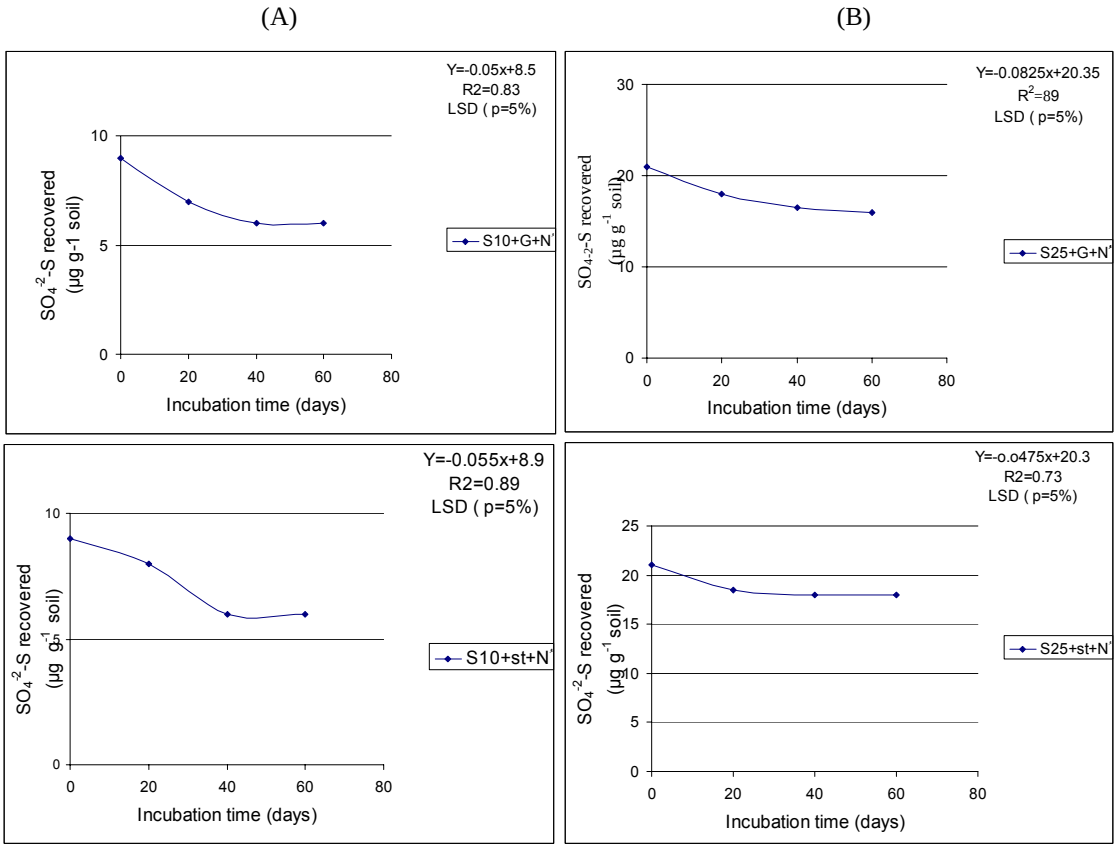


Fig. 2: Recovery of added SO_4^{-2}-S (A) S_{10} + G + N and S_{10} St + N treatments (B) S_{25} + G + N and S_{25} + St + N treatments. Details of the treatments are described in Table II

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