

# DNA Computing for an Absolute 1-Center Problem: An Evolutionary Approach

Zuwairie Ibrahim, Yusei Tsuboi, Osamu Ono, Marzuki Khalid

**Abstract**—Deoxyribonucleic Acid or DNA computing has emerged as an interdisciplinary field that draws together chemistry, molecular biology, computer science and mathematics. Thus, in this paper, the possibility of DNA-based computing to solve an absolute 1-center problem by molecular manipulations is presented. This is truly the first attempt to solve such a problem by DNA-based computing approach. Since, part of the procedures involve with shortest path computation, research works on DNA computing for shortest path Traveling Salesman Problem, in short, TSP are reviewed. These approaches are studied and only the appropriate one is adapted in designing the computation procedures. This DNA-based computation is designed in such a way that every path is encoded by oligonucleotides and the path's length is directly proportional to the length of oligonucleotides. Using these properties, gel electrophoresis is performed in order to separate the respective DNA molecules according to their length. One expectation arise from this paper is that it is possible to verify the instance absolute 1-center problem using DNA computing by laboratory experiments.

**Keywords**—DNA computing, operation research, 1-center problem.

## I. INTRODUCTION

IN 1965, Gordon E. Moore [1] observed an exponential growth in the number of transistor per integrated circuit against time. Currently, this is the definition of Moore's Law, meaning that more and more transistors can be crammed into a chip until the silicon itself reaches its finite atomic scale imitation. Since the traditional silicon-based computer is restricted by its fundamental physical limitation, researchers have been searching for alternative medium for computation and DNA would turn out to be the answer.

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Zuwairie Ibrahim was with the Department of Microelectronics and Computer Engineering, Faculty of Electrical Engineering, Universiti Teknologi Malaysia, 81310 Skudai, Johor Darul Takzim, Malaysia. He is now pursuing his PhD at the Institute of Applied DNA Computing, Meiji University, Kanagawa, 214-8571 Japan (phone: +81-44-934-7377; fax: +81-44-934-7909; e-mail: zuwairie@isc.meiji.ac.jp).

Yusei Tsuboi is with the Institute of Applied DNA Computing, Meiji University, Kanagawa, 214-8571 Japan (phone: +81-44-934-7377; fax: +81-44-934-7909; e-mail: tsuboi@isc.meiji.ac.jp).

Osamu Ono is the Director of Institute of Applied DNA Computing, Meiji University, Kanagawa, 214-8571 Japan (phone: +81-44-934-7377; fax: +81-44-934-7909; e-mail: ono@isc.meiji.ac.jp).

Marzuki Khalid is the Director of Center for Artificial Intelligence and Robotics (CAIRO), University Technology of Malaysia, Jalan Semarak, 54100 Kuala Lumpur, Malaysia (phone: +6-03-2691-3710; fax: +6-03-2697-0815; e-mail: marzuki@utmkl.utm.my).

A new computing paradigm based on DNA molecules is appeared in 1994 when Leonard M. Adleman [2] launched a novel approach to solve the so-called Hamiltonian Path Problem (HPP) with seven vertices by DNA molecules. While in conventional silicon-based computer, information is stored as binary numbers in silicon-based memory, he encoded the information of the vertices by a randomly DNA sequences. The computation is performed in biochemical reaction fashion involving hybridization, denaturation, ligation, Polymerase Chain Reaction (PCR), etc. The output of the computation, also in the form of DNA molecules can be read and printed by electrophoretical fluorescent method.

DNA molecules are composed of single or double DNA fragments or often called oligonucleotides or strands. Nucleotides form the basis of DNA. A single-stranded fragment has a phospho-sugar backbone and four kinds of bases denoted by the symbols A, T, G, and C for the bases adenine, thymine, guanine, and cytosine respectively. These four nucleic acids, which can occur in any order combined in Watson-Crick complementary pairs to form a double strand helix of DNA. Due to the hybridization reaction, A is complementary with T and C is complementary with G. As an example, any sequence oligonucleotides, such as 5'-ACCTG-3' has a complementary sequence, 3'-TGGAC-5'. Digits 5' and 3' denote orientation of DNA oligonucleotides.

The facility location optimization has many applications reported to date mostly in operations research and network design problems. It has been used in placement of routes [3] and internet web proxies [4], agglomeration of traffic [5-6], and web server replications in a content distribution network [7]. It can be largely divided into two main problems, namely MINSUM and MINMAX. MINSUM problem is to minimize the sum of the weighted distances to the nearest source facility. An example is p-medium problem [8]. On the other hand, MINMAX problem is to minimize the maximum weighted distance from the demand destinations to the nearest source facility. It is also called p-center problems [9]. Even if the number  $p=1$  but the network is large, the problems are almost NP-complete. It is then become relevant to develop a new method based on DNA computing where polynomial-time computation may be developed. For these reasons, the authors concentrate on solving an absolute 1-center problem using DNA-based computing.

Gigantic memory capacity and massively parallelism inherent in DNA computing can be exploited to make a brute force search on a big problem space in constant or polynomial-time. That is why DNA computing is well known very suitable for NP-complete problems. After Adleman's

novel discovery is published, various combinatorial problems have been chosen to express the possibility of DNA computing in many applications. It is reported that DNA computing is appropriate to compute the maximal clique problem [10], to solve satisfiability (SAT) problem [11] and even to break the Discrete Encryption Standard (DES) [12]. However, one of the combinatorial problems not being solved by DNA computing yet is the facility location optimization, in particular, an absolute 1-center problem. Nevertheless, this problem has been solved extensively by non-molecular or mathematical approaches and performed on sequential silicon based computer as computing platform. Some examples are greedy method [13], approximation algorithms [14] and by solving a series of set covering problems [15].

So far, nobody has proposed a molecular algorithm for performing 1-center problem. Hence, the authors confidently claim that the proposed approach is the first effort to solve the problem by DNA computing. In this paper, a new computational design is developed to solve an absolute 1-center problem by DNA-based computing approach. This idea is motivated from the previous research works carried out to compute the shortest path of TSP. The procedures are adapted and the overall design are modified and enhanced in order to solve the selected problem. The proposed computation necessarily composed of combination of DNA bio-chemical operation such as DNA synthesis, PCR, ligation, gel electrophoresis, annealing, melting and hybridization. More importantly, the proposed DNA-based computational approach may be easier to accomplish because the tedious affinity-purification operation will be never been used during the computation.

## II. MOLECULAR COMPUTATION FOR SHORTEST PATH PROBLEM

At first, Adleman pioneered the molecular computation by solving seven nodes of HPP. The goal of HPP is to determine whether a path exists that will commence at the start city, finish at the end city and pass through each city of the remaining cities exactly once. While this solution was very promising, the problem did not involve shortest path computation at all.

A variation of HPP, shortest path TSP tackled by Narayanan and Zorbalas [16] to solve by DNA algorithms. A constant increase of DNA strands according of the actual length of distance is occupied. A drawback of this method is that for some concatenations, concatenated DNA strand of two distances, while less numerically than the longest distance, may be longer than the DNA strands for that longest distance. This may leads to errors in computing shortest path. This scheme however was not been realized by laboratory experiments.

Yamamoto et al. presented concentration-controlled DNA computing for accomplishing a local search for shortest path problem [17]. Although DNA computing with concentration control method enables local search among all the candidate solutions, it is not guaranteed that the most intensive band is the DNA representing the shortest path in the given graph. In addition, it is technically difficult to extract a single optimal

solution from the most intensive band.

J. Y. Lee and his colleagues proposed temperature gradient-based DNA computing for shortest path TSP problem [18]. They introduce melting temperature ( $T_m$ ) for this purpose. At certain temperature, DNA strands of correct solutions will be denatured at that temperature, and be amplified by PCR process. As the denaturation temperature increases, other DNA strands also will be amplified. But the amount of correct solutions will be exponentially increased cycle by cycle, and occupy the major part of the solution. However, after hybridization, the temperature for the effective PCR process is not provided.

It is better to note that the absolute 1-center problem, even it involved shortest path computation as part of the problem, it requires computing the longest path among a set of shortest paths. It also requires computing the shortest path among a set of longest paths. Thus, the authors possibly found a molecular computation approach to do such computation. The computing strategy is motivated from [16] but we did a lot of modifications in order to make it suitable for computational needs. In this approach, the edge weights are handled by the DNA strands and the actual weights are approximated by the directly proportional length DNA strands. Hence, the important information during the computation is DNA length. The computation only considers about the starting node and end node encoded by DNA strands. The actual nodes encoded within the DNA strands will not be computed. As a result, the affinity-purification process is not required during the computation.

## III. PROBLEM FORMULATION

The 1-center problem may be phrased as follows. As given by Cheriyan and Ravi [19], the problem is to locate a facility on the points of a given network or graph so as to minimize the maximum cost of a path between a demand node and its nearest facility. A weighted undirected network or graph  $G = (V, E)$  with 5 vertices, 7 edges and 7 weights is used to model the problem where  $V$  and  $E$  are the vertices and edges of the graph respectively. If a path exists between the two nodes, a positive weight is assigned to that path. The graph is depicted in Fig. 1 [20].

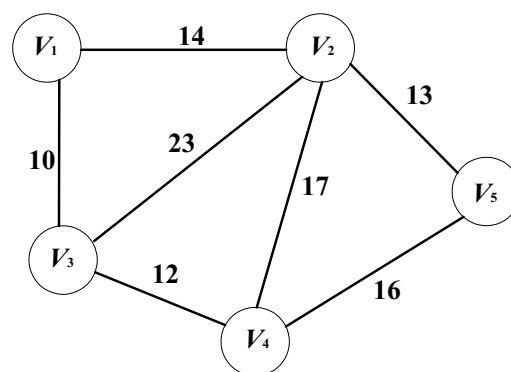


Figure 1: A weighted undirected graph  $G = (V, E)$ .

Normally, in order to solve this problem, one can note the shortest path between a particular node to its nearest node by a matrix as in Table 1 and then choose a node such that the maximum entry in its row in the matrix is smallest among the maximum entries of all rows. The underlined bold numbers highlights the maximum entries for each row. According to Table 1, it is clear that the right answer of this problem is to locate at node  $V_4$  with a maximum distance of 22.

TABLE I  
 NODE-TO-NODE DISTANCE BY A MATRIX

	$V_1$	$V_2$	$V_3$	$V_4$	$V_5$
$V_1$	0	14	10	22	<b><u>27</u></b>
$V_2$	14	0	<b><u>23</u></b>	17	13
$V_3$	10	23	0	12	<b><u>28</u></b>
$V_4$	<b><u>22</u></b>	17	12	0	16
$V_5$	27	13	<b><u>28</u></b>	16	0

#### IV. DNA BIOCHEMICAL OPERATIONS

There are a number of feasible DNA bio-chemical operations in DNA computing. As such, one can manipulate or generate a solution containing DNA strands by DNA synthesis, polymerase chain reaction (PCR), ligation and gel electrophoresis. In this section, these essential tools are described in more details as follows:

##### A. DNA Synthesis

At present, it is possible to get a test tube containing approximately  $10^{18}$  DNA molecules with a desired sequence. Some commercial DNA synthesis companies are available, which provide a reasonable price for this reason. As in 1998, Adleman [21] said that the cost for sequences of length 20 is just about \$25.

##### B. Denaturation

Double stranded DNA molecules can be separated without breaking the single strands by applying heat to the solution at about  $95^\circ\text{C}$ . The double stranded molecules will come apart because the hydrogen bonds between complementary nucleotides are much weaker than the covalent bond between adjacent nucleotides in the same strands.

##### C. Annealing, Concatenation, or Hybridization

As oppose to denaturation, by cooling down a DNA solution from approximately  $95^\circ\text{C}$  to  $55^\circ\text{C}$ , single Watson-Crick complementary DNA sequences will combined to form double stranded molecules. Also, based on this behavior, concatenation of two identical strands would be happened with the presence of a connective strand as depicted in Fig. 2. Concatenation is crucial in Adleman's demonstration and in this proposed approach as well.

##### D. Polymerase Chain Reaction

PCR is an incredible sensitive copying machine for DNA. It also can be used for DNA detection. Given a site-specific single molecule DNA, a million or even billion of similar molecules can be created by PCR process. In  $n$  steps, it can produces  $2^n$

copies of the same molecules. PCR needs a number of sub-sequence strands called 'primers', which is usually about 20 base long to signal a specific start and end site at a template for replication. PCR normally runs for 20-30 cycles of 3 steps as shown in Fig. 3. Each cycle consist of three phases: separating base pair strands of DNA at about  $95^\circ\text{C}$ , annealing at  $55^\circ\text{C}$  and extension at  $75^\circ\text{C}$  [22]. It takes about 3 hours normally to complete the cycles.

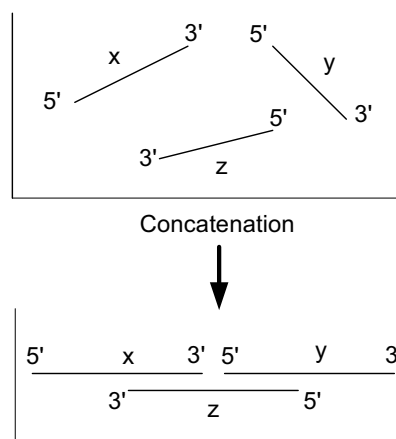


Figure 2: Concatenation of strand  $5' - x - 3'$  and  $5' - y - 3'$  with a presence of connective strand  $3' - z - 5'$ . If  $5' - x - 3' = 5' - x_1 x_2 - 3'$ ,  $5' - y - 3' = 5' - y_1 y_2 - 3'$ , then  $3' - z - 5' = 3' - \bar{x}_2 \bar{y}_1 - 5'$ , where  $\bar{x}_2$  and  $\bar{y}_1$  indicate the complementary strand of  $x_2$  and  $y_1$  respectively.

##### E. Ligation

Ligation often invoked after the single DNA strands are annealed and concatenated to each other. Many single-strand fragments will be connected in series and ligase is used as 'glue' to seal the covalent bonds between the adjacent fragments as shown in Figure 4 [23].

##### F. Gel Electrophoresis

DNA strands in a solution can be separated in term of its length by means of gel electrophoresis. In fact, the molecules are separated according to their weight, which is almost proportional to their length [24]. This technique is based on the fact that DNA molecules are negatively charged [25]. Hence, by putting them in an electric field, they will move towards the positive electrode at different speed. The longer molecules will remain behind the shorter ones. The speed also depends on the gel porosity. Agarose gel is frequently used and by varying the porosity of the gel used, the sensitivity of this length separation operation can be altered. The precision is high even molecules which differ by one nucleotide can still be distinguished between each other. An example of gel electrophoresis process [26] and its output [27] are well depicted in Figure 5 and Figure 6 respectively. This technique can be used to "print" the results of DNA computation as well. Normally, at the end of this process, the gel is photographed for convenience.

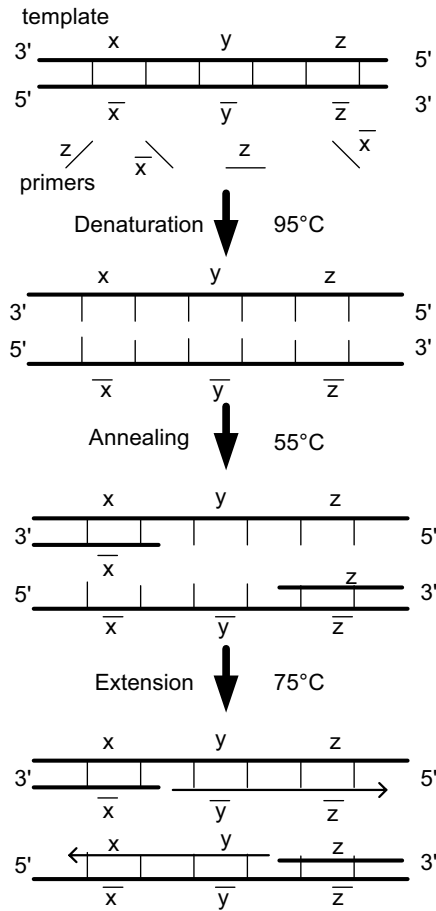


Figure 3: One PCR cycle. For 30 cycles of PCR, these 3 steps are repeated for 30 times.

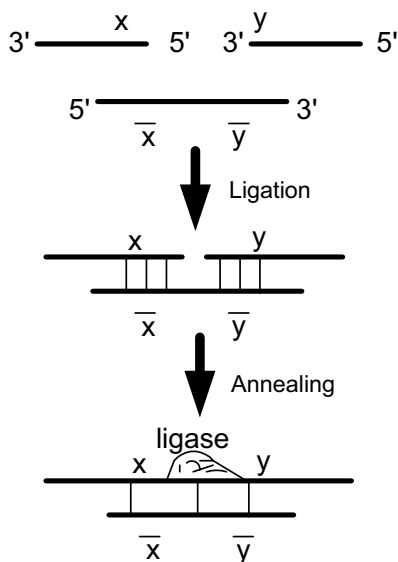


Figure 4: Ligation reaction.

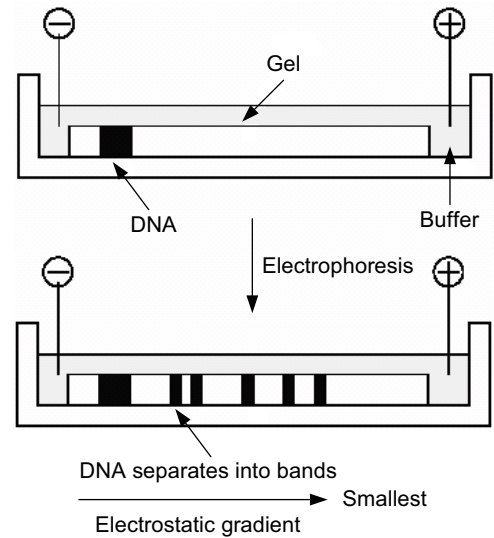


Figure 5: Gel electrophoresis process.

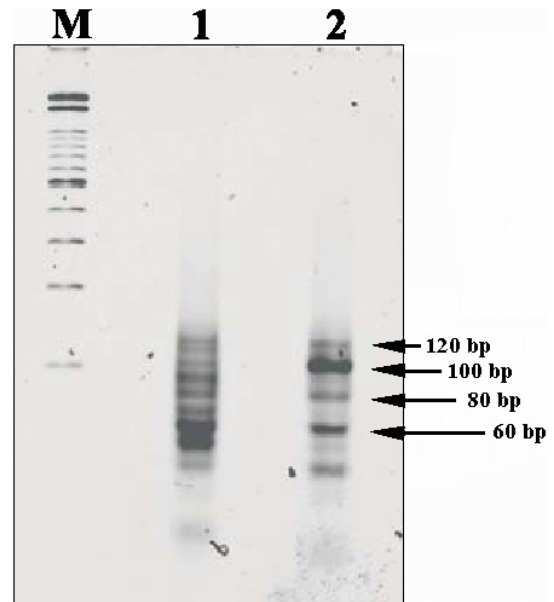


Figure 6: Gel electrophoresis output where lane M is DNA size marker. Lane 1 and 2 are used for the tested DNA molecules.

## V. COMPUTING WITH DNA

The overall computation described in this section consists of five steps altogether. Step 1 through step 3 is crucial for the generating initial pool, random route formation, whereas step 4 through step 6 is designed for the computation process and applied to the initial pool of solution.

### A. Step 1

Let  $V$  be the total number of nodes in the graph and  $E$  the total number of paths in the graph. The random sequence strands correspond to all nodes (and its complements) and distances (and its complements) are created. Let  $O_i$  ( $i = 1, \dots,$

$V$ ) and  $\overline{O}_i$  ( $i = 1, \dots, V$ ) be the fixed length random sequences correspond to all nodes in the graph and its complements respectively. In the same way, let  $O_d$  ( $d = 1, \dots, E$ ) and  $\overline{O}_d$  ( $d = 1, \dots, E$ ) be the variable length random sequences correspond to all the distances and its complements respectively. The length  $l$  of  $O_d$  and  $\overline{O}_d$  will be directly proportional to the weight distances. For instance, let say, for a constant proportional increase of 2, all the random sequences are placed in Table 2 and Table 3. For the sake of convenience, in Table 2 and Table 3, in order to avoid clumsy presentation, we will use symbolic representation of the distance sequences. Recall that these sequences would be occurred in any order of nucleotides and its length will be directly proportional to the distances. At the end of this step, oligonucleotides  $\overline{O}_d$  and  $\overline{O}_i$  are synthesized.

TABLE II  
 FIXED LENGTH NODE RANDOM SEQUENCES

Node	DNA Code (5'-3')	Complements (3'-5')
$V_1$	TATT	ATAA
$V_2$	GCGG	CGCC
$V_3$	GTCG	CAGC
$V_4$	AGAA	TCTT
$V_5$	TCCG	AGGC

TABLE III  
 VARIABLE LENGTH DISTANCE RANDOM SEQUENCES

Edge Cost	DNA Code (5'-3')	Complements (3'-5')
$D \{10\}$	20	20
$D \{12\}$	24	24
$D \{13\}$	26	26
$D \{14\}$	28	28
$D \{16\}$	32	32
$D \{17\}$	34	34
$D \{23\}$	46	46

B. Step 2

Let  $i$  be the start node of a path,  $d$  the path's distance and  $j$  the end node of that path. Oligonucleotides representing every path between two nodes in the graph are synthesized as follows:

synthesize oligonucleotides

$O_{i-d-j\_end}$  as **HR**  $O_i$  + **ALL**  $O_d$  + **ALL**  $O_j$

$O_{j-d-i\_end}$  as **HR**  $O_j$  + **ALL**  $O_d$  + **ALL**  $O_i$

$O_{i-d-j}$  as **HR**  $O_i$  + **ALL**  $O_d$  + **HL**  $O_j$

$O_{j-d-i}$  as **HR**  $O_j$  + **ALL**  $O_d$  + **HL**  $O_i$

where '**ALL**' represents the whole DNA strand, '**HL**' the left

half part, '**HR**' the right half part and '+' is a join. For each path, connecting two identical nodes, each of the nodes could be a starting node, an end node or a connective node in a path. Thus, four identical oligonucleotides will be created for each edge.

As such, if  $O_i = \text{TATT}$  then  $\overline{O}_i = \text{ATAA}$ ; if  $O_j = \text{GCGG}$  then  $\overline{O}_j = \text{CGCC}$  and if  $O_d = \text{ACGACGTT}$  then  $\overline{O}_d = \text{TCGTGCAA}$ . Then the path  $O_{i-d-j\_end}$  is **TTACGACGTTGCGG** (**HR**  $O_i$  + **ALL**  $O_d$  + **ALL**  $O_j$ ), the path  $O_{j-d-i\_end}$  is **GGACGACGTTTATT** (**HR**  $O_j$  + **ALL**  $O_d$  + **ALL**  $O_i$ ), the path  $O_{i-d-j}$  is **TTACGACGTTGC** (**HR**  $O_i$  + **ALL**  $O_d$  + **HL**  $O_j$ ), and lastly the path  $O_{j-d-i}$  is **GGACGACGTTTA** (**HR**  $O_j$  + **ALL**  $O_d$  + **HL**  $O_i$ ).

At the end of this step, 28 identical edge strands will be created by a combination of the nodes sequences and distance sequences. Because of this combination, the edges strands  $O_{i-d-j}$ ,  $O_{i-d-j\_end}$ ,  $O_{j-d-i}$ , and  $O_{j-d-i\_end}$  listed in Table 4 are influenced by the additional nodes bases and hence, unable to handle the weight of each edges. The additional bases would cause errors during the computation because the additional bases composed in the edge sequences reflect the weight value of edges. To compensate the additional bases, the strands corresponds to distance sequences are shorten according to the additional bases and the node sequences in the edge strands are left unchanged. All the modified oligonucleotides  $O_{i-d-j(modified)}$ ,  $O_{i-d-j\_end(modified)}$ ,  $O_{j-d-i(modified)}$ , and  $O_{j-d-i\_end(modified)}$  are listed in Table 5. This modification however, decreases the generality of the approach because the weight of edges less than 3 is not allowed.

C. Step 3

All modified oligonucleotides  $O_{i-d-j(modified)}$ ,  $O_{i-d-j\_end(modified)}$ ,  $O_{j-d-i(modified)}$ , and  $O_{j-d-i\_end(modified)}$  followed by all complement oligonucleotides ( $\overline{O}_i$  and  $\overline{O}_d$ ) are inserted into a test tube and DNA ligase reaction is performed in which random routes through the graph are formed. In this way, the initial solution,  $N_0$ , which contains a large set of random routes of the graph are generated. After that, in order to make sure that no sticky end strands exist in the solution, polymerase enzyme is used for the extension where the sticky end strands are altered to become blunt end strands.

At this moment, an initial solution  $N_0$  is already obtained and it is time to filter out the right routes among the vast random routes of the graph. Unlike the conventional filtering, it is not merely throwing away the unwanted strands but rather copying the right strands exponentially by incredibly sensitive PCR process.

D. Step 4

Produce another five solutions from the initial solution,  $N_0$ . Each solution represents exactly one row in Table 1.

TABLE IV  
 EDGE SEQUENCES

Edges	DNA Code (5'-3')
$V_3 - V_1$	CG <span style="border: 1px solid black; padding: 2px;">20</span> TATT
	TT <span style="border: 1px solid black; padding: 2px;">20</span> GTCG
	CG <span style="border: 1px solid black; padding: 2px;">20</span> TA
	TT <span style="border: 1px solid black; padding: 2px;">20</span> GT
$V_4 - V_3$	AA <span style="border: 1px solid black; padding: 2px;">24</span> GTCG
	CG <span style="border: 1px solid black; padding: 2px;">24</span> AGAA
	AA <span style="border: 1px solid black; padding: 2px;">24</span> GT
	CG <span style="border: 1px solid black; padding: 2px;">24</span> AG
$V_2 - V_5$	GG <span style="border: 1px solid black; padding: 2px;">26</span> TCCG
	CG <span style="border: 1px solid black; padding: 2px;">26</span> GCGG
	GG <span style="border: 1px solid black; padding: 2px;">26</span> TC
	CG <span style="border: 1px solid black; padding: 2px;">26</span> GC
$V_1 - V_2$	TT <span style="border: 1px solid black; padding: 2px;">28</span> GCGG
	GG <span style="border: 1px solid black; padding: 2px;">28</span> TATT
	TT <span style="border: 1px solid black; padding: 2px;">28</span> GC
	GG <span style="border: 1px solid black; padding: 2px;">28</span> TA
$V_5 - V_4$	CG <span style="border: 1px solid black; padding: 2px;">32</span> AGAA
	AA <span style="border: 1px solid black; padding: 2px;">32</span> TCCG
	CG <span style="border: 1px solid black; padding: 2px;">32</span> AG
	AA <span style="border: 1px solid black; padding: 2px;">32</span> TC
$V_4 - V_2$	AA <span style="border: 1px solid black; padding: 2px;">34</span> GCGG
	GG <span style="border: 1px solid black; padding: 2px;">34</span> AGAA
	AA <span style="border: 1px solid black; padding: 2px;">34</span> GC
	GG <span style="border: 1px solid black; padding: 2px;">34</span> AG
$V_2 - V_3$	GG <span style="border: 1px solid black; padding: 2px;">46</span> GTCG
	CG <span style="border: 1px solid black; padding: 2px;">46</span> GCGG
	GG <span style="border: 1px solid black; padding: 2px;">46</span> GT
	CG <span style="border: 1px solid black; padding: 2px;">46</span> GC

TABLE V  
 MODIFIED EDGE SEQUENCES

Edge	DNA Code (5'-3')
$V_3 - V_1$	CG <span style="border: 1px solid black; padding: 2px;">14</span> TATT
	TT <span style="border: 1px solid black; padding: 2px;">14</span> GTCG
	CG <span style="border: 1px solid black; padding: 2px;">16</span> TA
	TT <span style="border: 1px solid black; padding: 2px;">16</span> GT
$V_4 - V_3$	AA <span style="border: 1px solid black; padding: 2px;">18</span> GTCG
	CG <span style="border: 1px solid black; padding: 2px;">18</span> AGAA
	AA <span style="border: 1px solid black; padding: 2px;">20</span> GT
	CG <span style="border: 1px solid black; padding: 2px;">20</span> AG
$V_2 - V_5$	GG <span style="border: 1px solid black; padding: 2px;">20</span> TCCG
	CG <span style="border: 1px solid black; padding: 2px;">20</span> GCGG
	GG <span style="border: 1px solid black; padding: 2px;">22</span> TC
	CG <span style="border: 1px solid black; padding: 2px;">22</span> GC
$V_1 - V_2$	TT <span style="border: 1px solid black; padding: 2px;">22</span> GCGG
	GG <span style="border: 1px solid black; padding: 2px;">22</span> TATT
	TT <span style="border: 1px solid black; padding: 2px;">24</span> GC
	GG <span style="border: 1px solid black; padding: 2px;">24</span> TA
$V_5 - V_4$	CG <span style="border: 1px solid black; padding: 2px;">26</span> AGAA
	AA <span style="border: 1px solid black; padding: 2px;">26</span> TCCG
	CG <span style="border: 1px solid black; padding: 2px;">28</span> AG
	AA <span style="border: 1px solid black; padding: 2px;">28</span> TC
$V_4 - V_2$	AA <span style="border: 1px solid black; padding: 2px;">28</span> GCGG
	GG <span style="border: 1px solid black; padding: 2px;">28</span> AGAA
	AA <span style="border: 1px solid black; padding: 2px;">30</span> GC
	GG <span style="border: 1px solid black; padding: 2px;">30</span> AG
$V_2 - V_3$	GG <span style="border: 1px solid black; padding: 2px;">40</span> GTCG
	CG <span style="border: 1px solid black; padding: 2px;">40</span> GCGG
	GG <span style="border: 1px solid black; padding: 2px;">42</span> GT
	CG <span style="border: 1px solid black; padding: 2px;">42</span> GC

E. Step 5

- For each solution (or each row);
- Select a particular node as a starting point and find the shortest path from the starting node to other nodes.
  - Store the strands representing each shortest path into a solution.
  - Gather all together the solution representing all shortest paths.
  - Search the longest strand among the shortest strands, which indicates the maximum path of a particular row in Table 1.

F. Step 6

Gather all together the solutions that represent all the longest paths of each row. Search the shortest strand. Find the starting node of that path to get the answer of the absolute 1-center problem. The overall procedures are summarized by a flow chart given in Fig. 7.

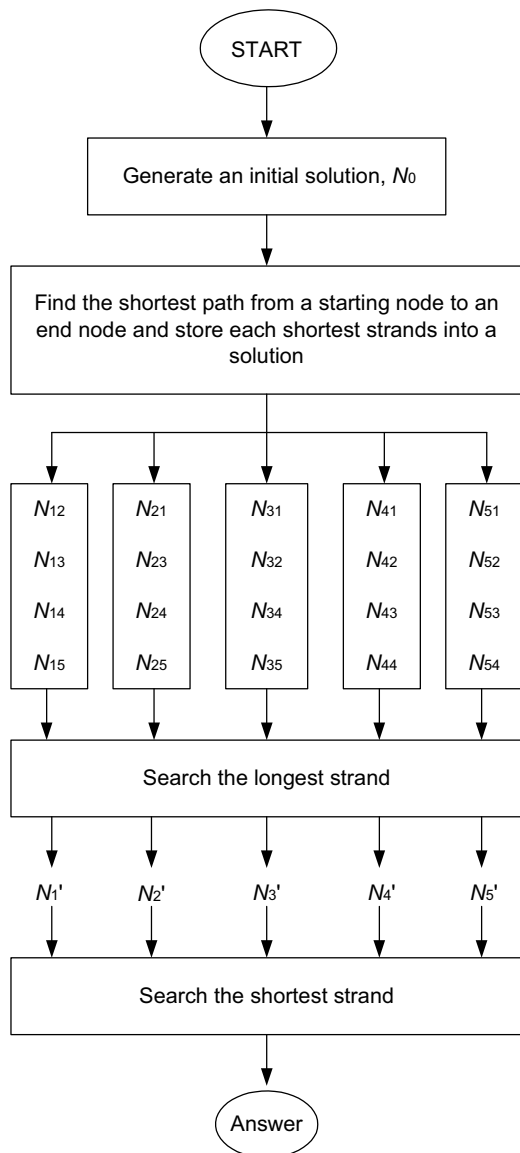


Figure 7: Overall procedures.

VI. DISCUSSIONS

The fourth step could be done by merely divides the solution  $N_0$  equally into five solutions and marks the solutions as  $N_1, N_2, N_3, N_4,$  and  $N_5$ . In this work, it is assumed that all the strands are spread to all areas in the solution.

It is possible to carry out the fifth step by the subsequent procedure. Since the dividing operation is a volume decreasing operation, it is important, therefore, to increase the volume of solutions to a sufficient amount. The only operation, which is able to increase the volume of the solutions, is the polymerase chain reaction (PCR) operation. Besides, this operation selects the routes that begin with a particular node for each test tube obtained in the previous step. Five types of primers will be employed where one primer, encoding the 3'-half of starting node is assigned to one particular test tube as shown in Fig. 8.

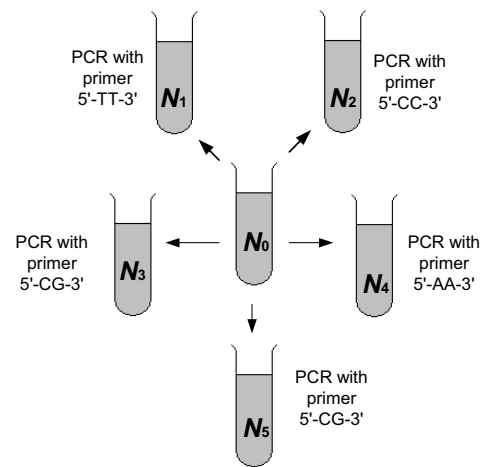


Figure 8: The initial solution  $N_0$  is divided equally into 5 solutions. PCR process is executed to each solution with 5 different primers representing 5 starting nodes.

Next, closely similar to the previous operation, by assuming that all the strands are spread uniformly in the solution, each solution obtained from previous operation,  $N_1, N_2, N_3, N_4,$  and  $N_5$  are divided equally into another 4 test tube. As a result, there should be 20 test tubes all together. Next, DNA strands with a particular end node are amplified and consequently, the volume of solutions is increase as well. PCR process is employed for each test tube by primers encoding the 3'-half of starting node and complement of the end nodes. This operation is well depicted in Fig. 9.

For better understanding, take a look closely at the molecules of solution  $N_{12}$  only. After two PCR operations are accomplished, there should be a numerous number of strands representing the beginning node,  $V_1$  and the end node,  $V_2$ . Some of them are shown in Fig. 10. Next, gel electrophoresis technique will separates these strands in term of length and the shortest length is chosen in order to obtain the shortest path. As a result, the strands  $V_1 \rightarrow V_2$  will be chosen. The output solution is called  $N_{12}'$ .

The same procedure will be executed to the solution  $N_{13}, N_{14},$

and  $N_{15}$  results in 3 solutions,  $N_{12}$ ,  $N_{13}$ , and  $N_{15}$  consisting the shortest strands  $V_1 \rightarrow V_3$ ,  $V_1 \rightarrow V_3 \rightarrow V_4$ , and  $V_1 \rightarrow V_2 \rightarrow V_5$  respectively. These strands are shown in Fig. 11.

All these four solutions,  $N_{12}$ ,  $N_{13}$ ,  $N_{14}$ , and  $N_{15}$  will be gathered together and due to the previous gel electrophoresis outputs, the longest strand will be selected among the shortest strands. Meaning that the strand  $N_{15}$  will be selected. The resultant solution will be called as  $N_1$ . The same procedures for  $N_1$  is repeated to the solution  $N_2$ ,  $N_3$ ,  $N_4$ , and  $N_5$  and the output solution  $N_2$ ,  $N_3$ ,  $N_4$ , and  $N_5$  respectively will be produced. At this stage, the sixth step is already completed.

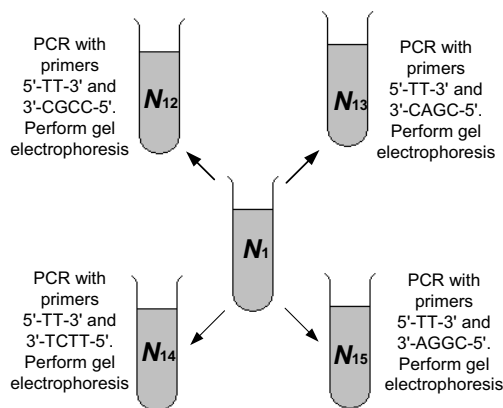


Figure 9: For instance,  $N_1$  is divided into another four test tubes namely  $N_{12}$ ,  $N_{13}$ ,  $N_{14}$ , and  $N_{15}$ . PCR process is executed again to each solution with primers representing the 3' half of start node and complement of end nodes. Next, for each test tube, the gel electrophoresis is applied for separating the strands by length.

TT	22	GCGG
AA	22	CGCC

(a)

TT	14	GT	CG	42	GCGG
AA	14	CA	GC	42	CGCC

(b)

TT	14	GT	CG	20	AG	AA	30	GCGG
AA	14	CA	GC	20	TC	TT	30	CGCC

(c)

Figure 10: (a) Strand  $V_1 \rightarrow V_2$  (b) Strand  $V_1 \rightarrow V_3 \rightarrow V_2$ , and (c) Strand  $V_1 \rightarrow V_3 \rightarrow V_4 \rightarrow V_2$ .

Lastly, the sixth step begins with collecting the solution  $N_1$ ,  $N_2$ ,  $N_3$ ,  $N_4$ , and  $N_5$ . According to the previous gel electrophoresis results, the solution representing the shortest strand will be chosen manually. This solution indicates the actual answer of the absolute 1-center problem. The selection involved based on gel electrophoresis is shown in Fig. 12.

TT	14	GTCG
AA	14	CAGC

(a)

TT	14	GT	CG	20	AGAA
AA	14	CA	GC	20	TCTT

(b)

TT	22	GC	GG	22	TCCG
AA	22	CG	CC	22	AGAA

(c)

Figure 11: (a) Strand  $V_1 \rightarrow V_3$  (b) Strand  $V_1 \rightarrow V_3 \rightarrow V_4$ , and (c) Strand  $V_1 \rightarrow V_2 \rightarrow V_5$ .

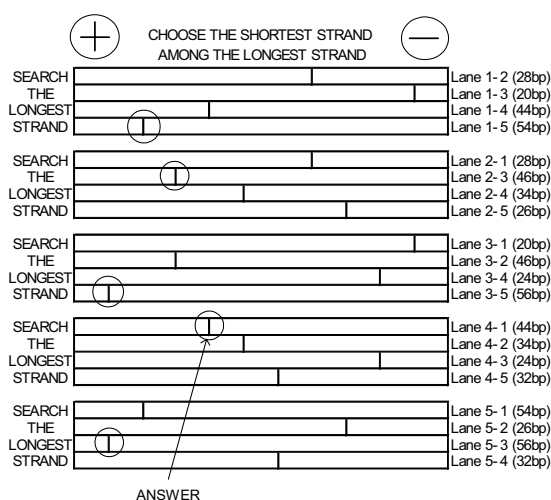


Figure 12: The selection based on expectation result of gel electrophoresis.

Heuristically, at this stage, there is only one type of strand exist. The DNA strands indicating the path from node  $V_4$  to  $V_1$  (or explicitly the path from node  $V_4$  to  $V_3$  to  $V_1$ ). This expected result shows that one has to locate the facility on point  $V_4$  in order to minimize the maximum cost of a path between a demand node to its nearest facility.

## VII. CONCLUSIONS

This objective of this paper is to certify the feasibility of DNA computing in an absolute 1-center problem. The authors carried out the first attempt to compute 1-center problem by DNA-based computing approach. The contents of this paper also imply a possibility to verify the proposed procedures by experimental work. Since the computing platform is now different, it is not relevant to make a comparison between the proposed approach and other existing approaches. Moreover, this computing platform is still infant compared to that matured conventional silicon-based computation. Despite all the



limitations, the usefulness and strong potentials of DNA-based computational approach provides a compelling reason to explore a lot of possible applications of DNA computing and solving an absolute 1-center problem is one of them.

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**Zuwairie Ibrahim** received the B.Eng (Mechatronics) and M.Eng. (Image Processing) from Universiti Teknologi Malaysia, Malaysia, in 2000 and 2002 respectively. Since 2002, he engaged with Department of Mechatronics and Robotics, Universiti Teknologi Malaysia as a lecturer. He is currently pursuing his PhD at the Institute of Applied DNA Computing, Meiji University, Kanagawa, Japan. He is a student member of Institute of Electrical and Electronics Engineers (IEEE), International Computational Intelligence Society (ICIS), and International Signal Processing Society (ISPC). His research interests include signal and image processing, automated visual inspection, evolutionary and unconventional computing such as molecular or DNA computing.

**Yusei Tsuboi** received the B. Eng and Master degree from Meiji University, Japan, in 2000 and 2002 respectively. Currently, he is pursuing the PhD degree also at Meiji University, Japan. Presently, his research interest includes robotics and DNA computing for artificial intelligence.

**Osamu Ono** received the Bachelor, Master and Doctor Degree in Engineering all from Waseda University, Tokyo, in 1974, 1976, and 1979 respectively. He is a Professor of the Department of Electrical and Electronic Engineering, Meiji University, Japan. He is the Director of Tokyo Branch of Japan Institute Electrical Engineering and a committee member of Japan Society for Simulation Technology (JSST). His research interest includes large scale industrial process, mechatronics, advanced mobile robotics and image processing. Currently, he is interested on the application of DNA computing in engineering field.

**Marzuki Khalid** is currently a Professor in Intelligent Control and Director at the Centre of Artificial Intelligence and Robotics (CAIRO) of Universiti Teknologi Malaysia. His current research interest is in the field of artificial intelligence with applications to control. He has been appointed to the Editorial Advisory Board of the International Journal of Engineering Applications of Artificial Intelligence published by Elsevier Science and an Associate Editor of the Journal of Systems and Control Engineering published by the Institute of Mechanical Engineers, United Kingdom. He is currently the IEEE Student Activities Chair for Region 10 (Asia Pacific) and also the Founding Member of the Asian Control Professors Association.